

# Pan-genomics: theory & practice

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Sept 20, 2014

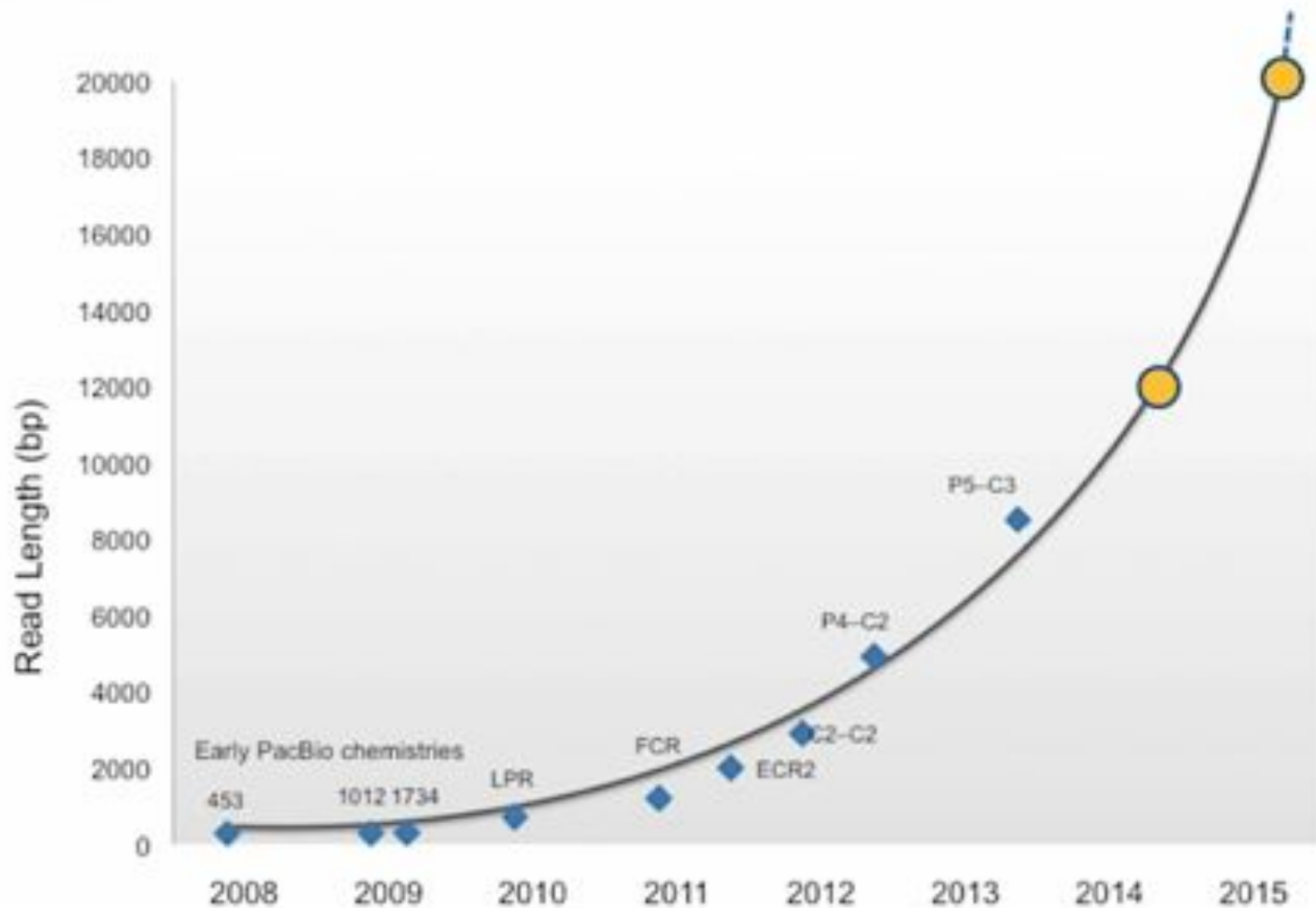
GRC Assembly Workshop



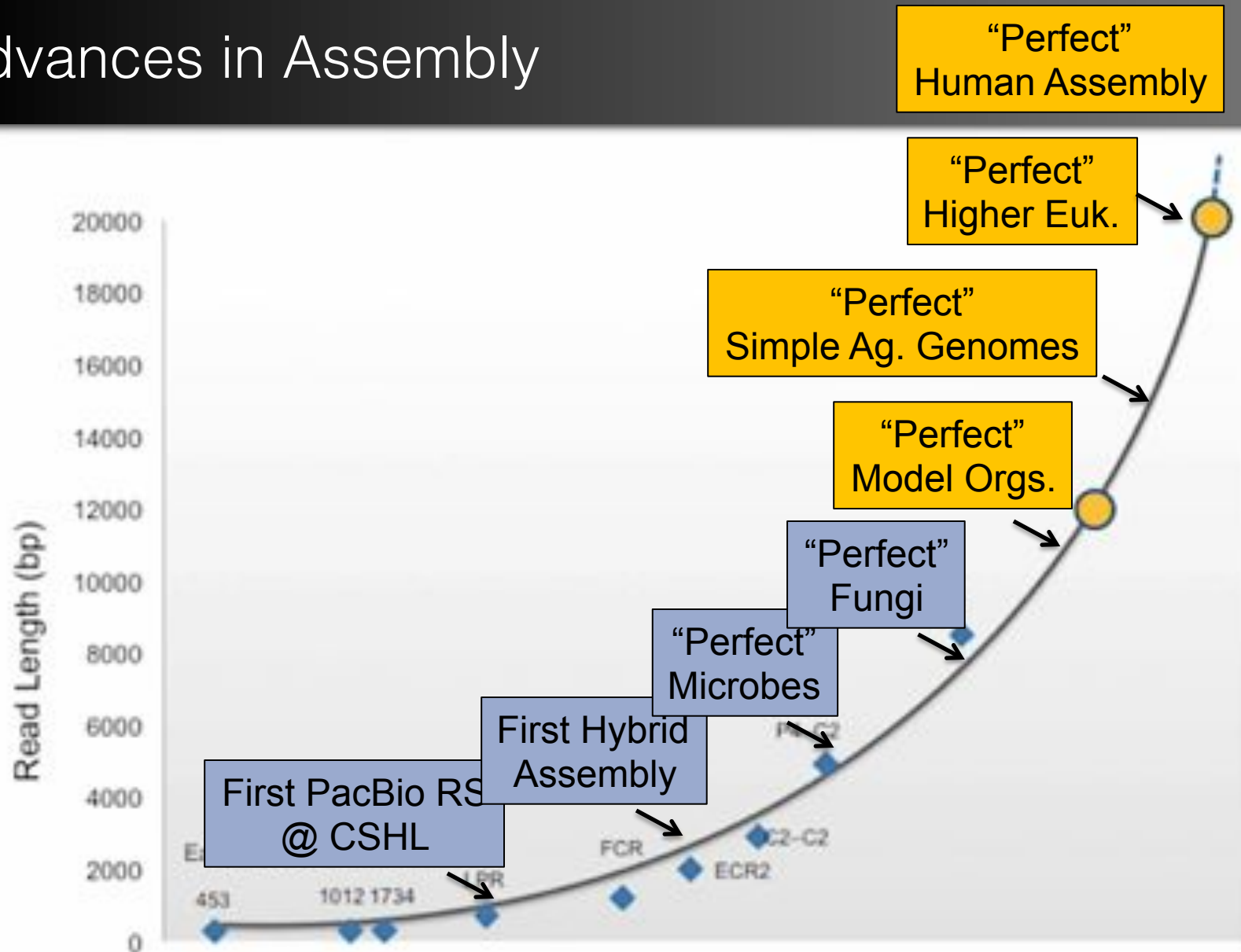
#gi2014 / @mike\_schatz

# Part I: Theory

# PacBio® Advances in Read Length



# Advances in Assembly

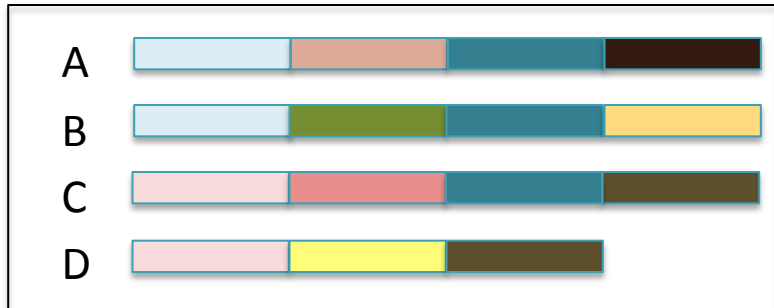


**Error correction and assembly complexity of single molecule sequencing reads.**

Lee, H\*, Gurtowski, J\*, Yoo, S, Marcus, S, McCombie, WR, Schatz, MC

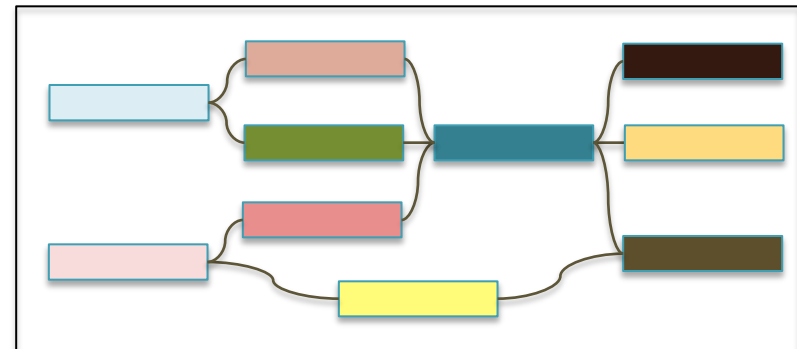
<http://www.biorxiv.org/content/early/2014/06/18/006395>

# Pan-Genome Alignment & Assembly



Time to start considering problems for which  $N$  complete genomes are the input to study the “pan-genome”

- Available today for many microbial species, near future for higher eukaryotes



Pan-genome colored de Bruijn graph

- Encodes all the sequence relationships between the genomes
- How well conserved is a given sequence?
- What are the pan-genome network properties?

## SplitMEM: Graphical pan-genome analysis with suffix skips

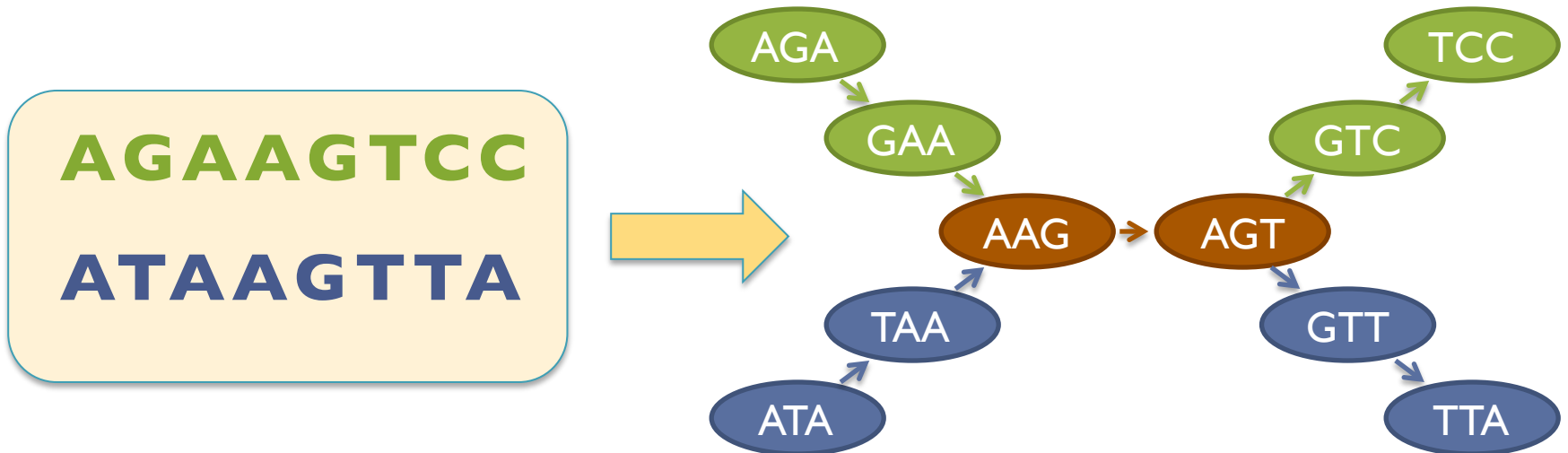
Marcus, S, Lee, H, Schatz, MC

<http://biorxiv.org/content/early/2014/04/06/003954>

# Graphical pan-genome analysis

## Colored de Bruijn graph

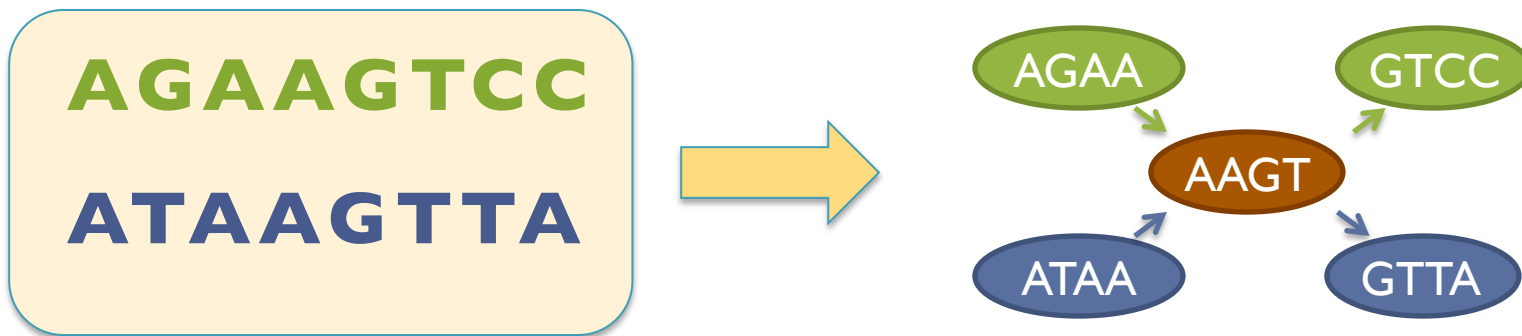
- Node for each distinct kmer
- Directed edge connects consecutive kmers
- Nodes overlap by  $k-1$  bp



# Graphical pan-genome analysis

## Colored de Bruijn graph

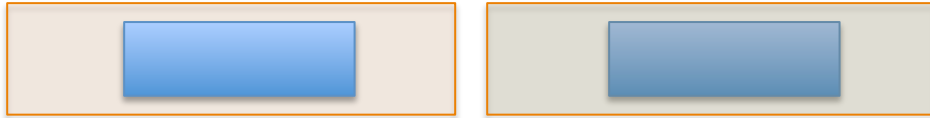
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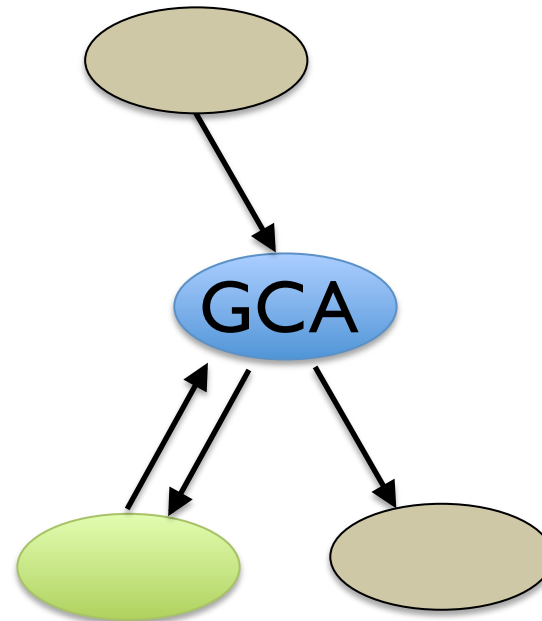
### **More specifically:**

- We aim to build the compressed de Bruijn graph as quickly as possible without considering every distinct kmer

# Maximal Exact Matches (MEMs) to de Bruijn Graphs

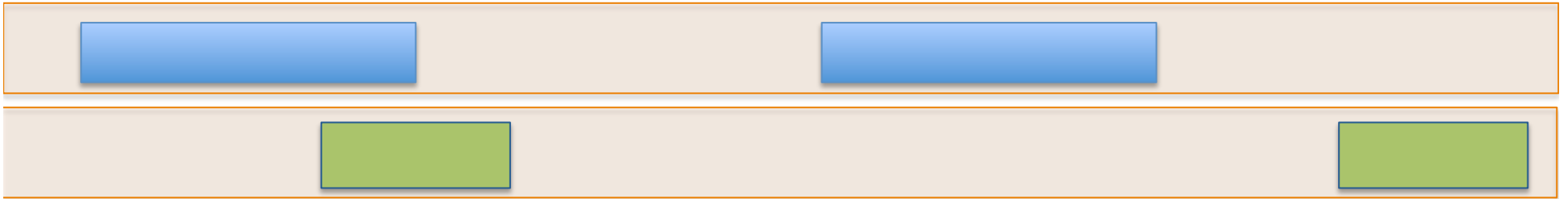


TGCAC...GGCAA

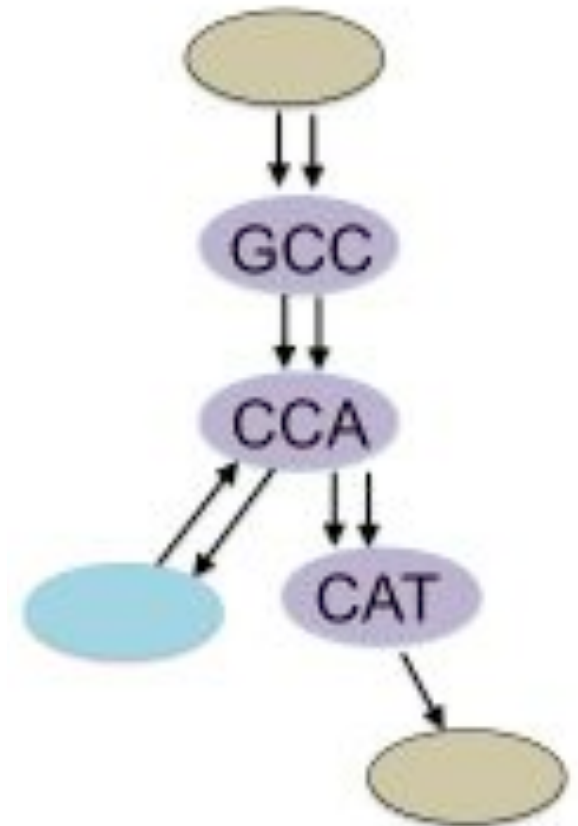




# Overlapping MEMs

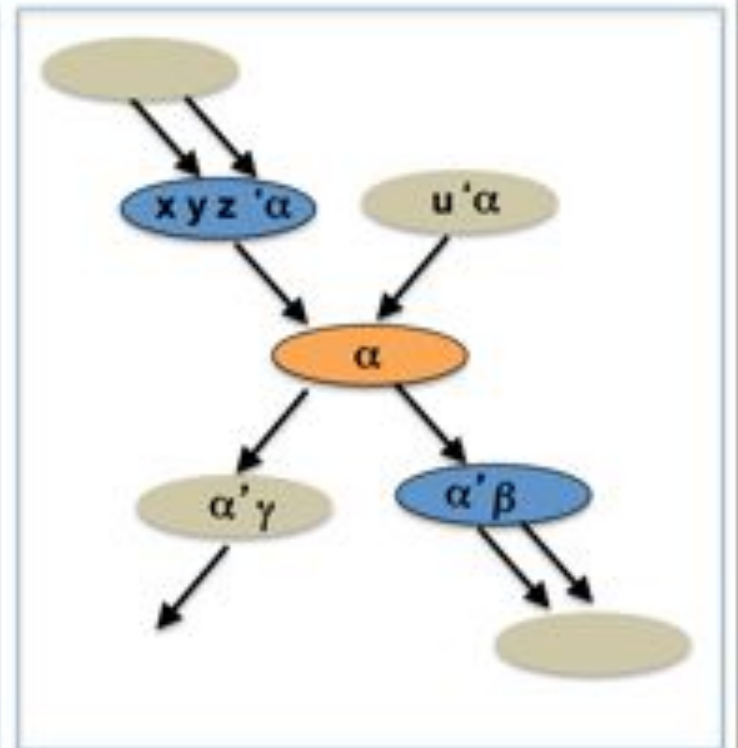
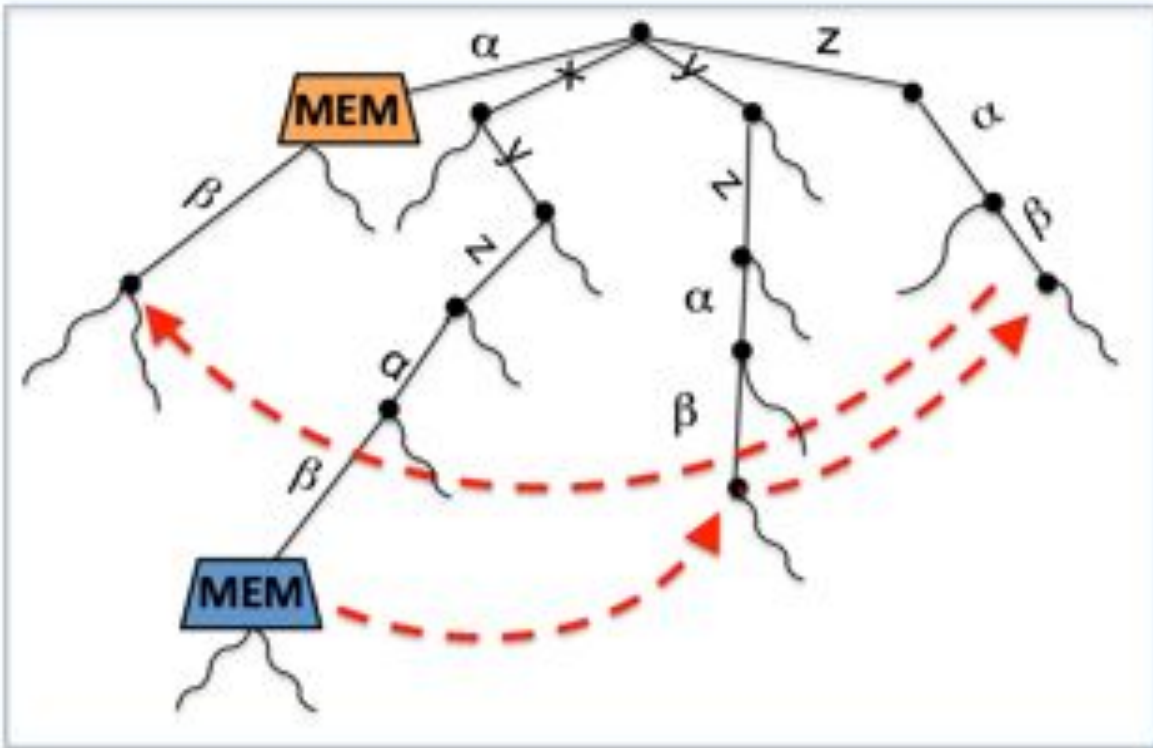


TGCCATCGCCAACCAT  
TGCCATCGCCAACCAT



# Suffix Trees & de Bruijn Graphs

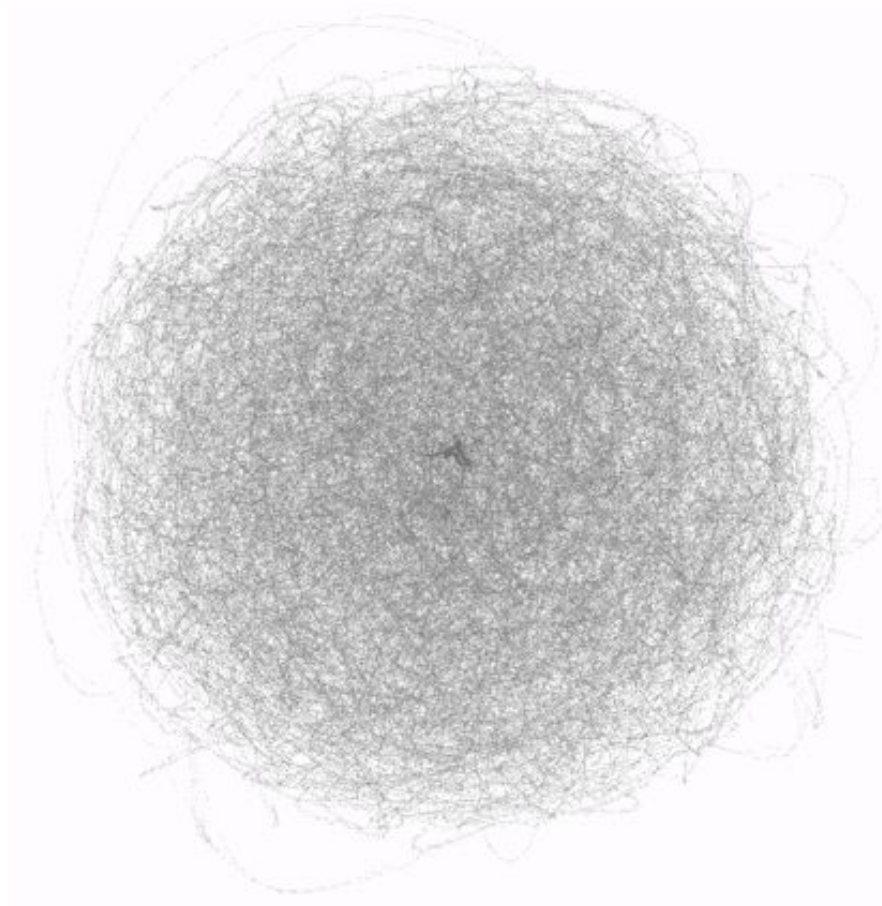
... x **xyz** **α** **β** ... y **xyz** **α** **β** ... u **α** **γ** ...



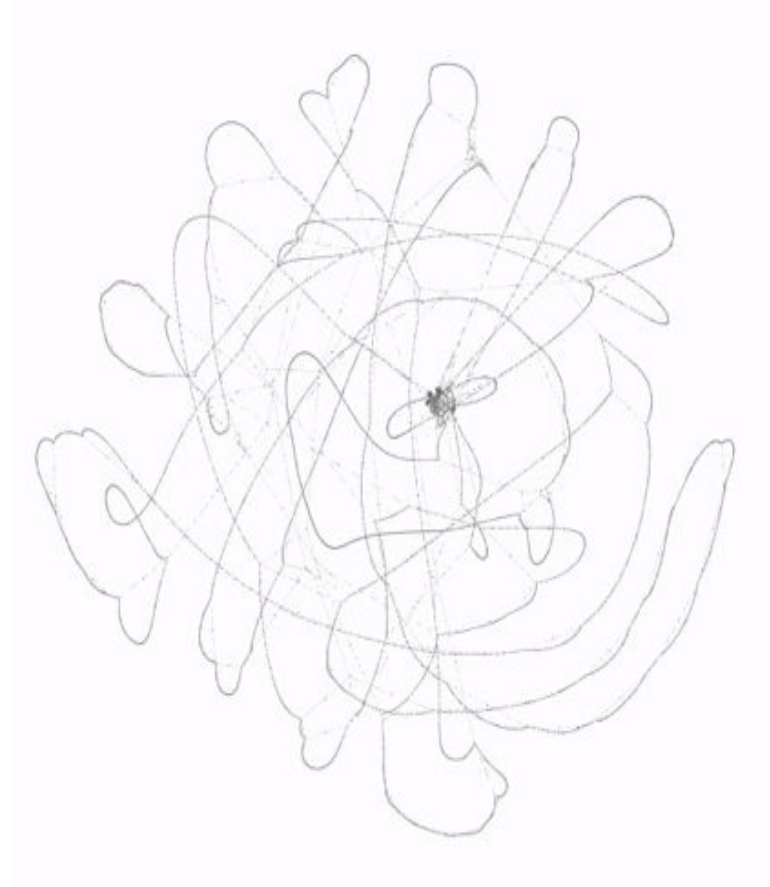
## Key concepts:

- Shared sequences form repeats called “maximal exact matches” (MEM)
- Easy to identify MEMs in a suffix tree, but may be nested within other MEMs
- Use “suffix skips” to quickly decompose MEMs, add in the missing nodes and edges

# *B. anthracis* pan-genome (9 strains)



k=25

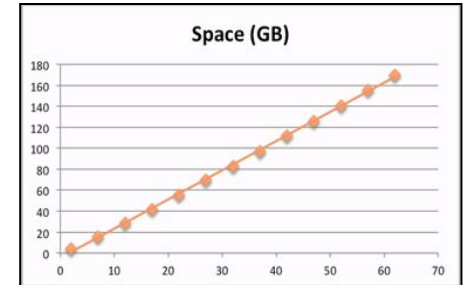


k=1000

# Microbial Pan-Genomes

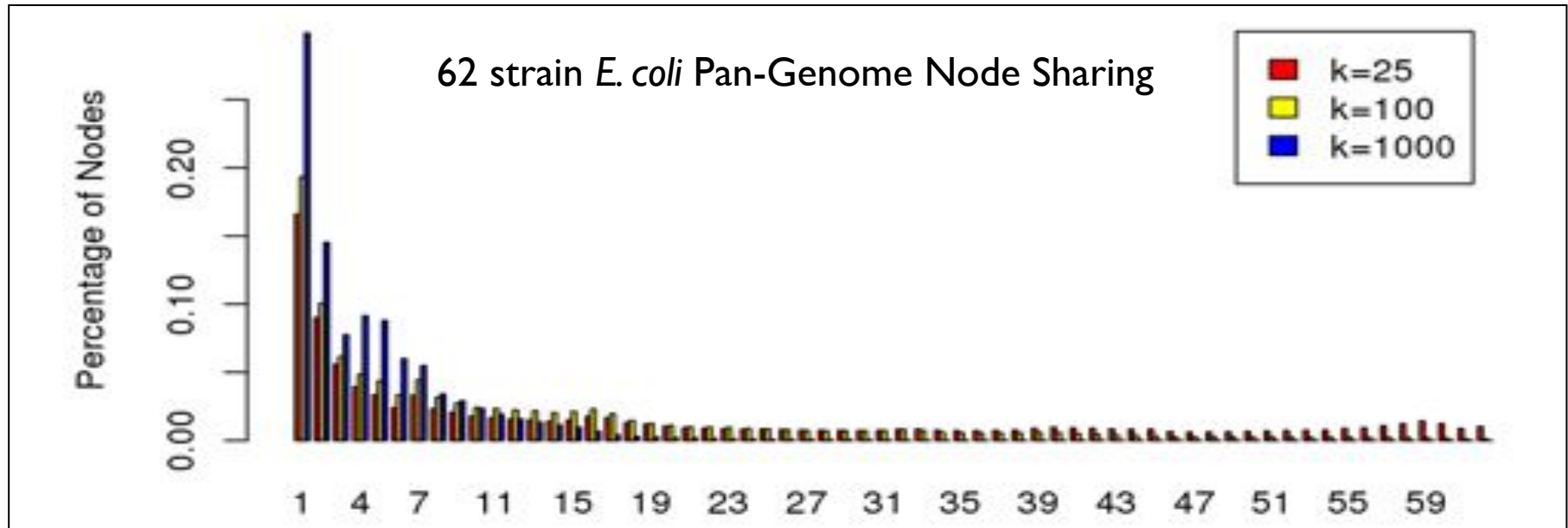
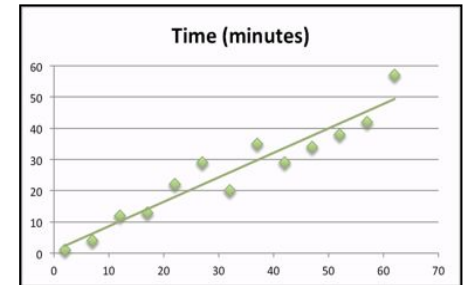
## **E. coli (62) and B. anthracis (9) pan-genome analysis**

- Analyzed all available strains in Genbank
- Space and time are effectively linear in the number of genomes
  - $O(n \log g)$  where  $g$  is the length of the longest genome



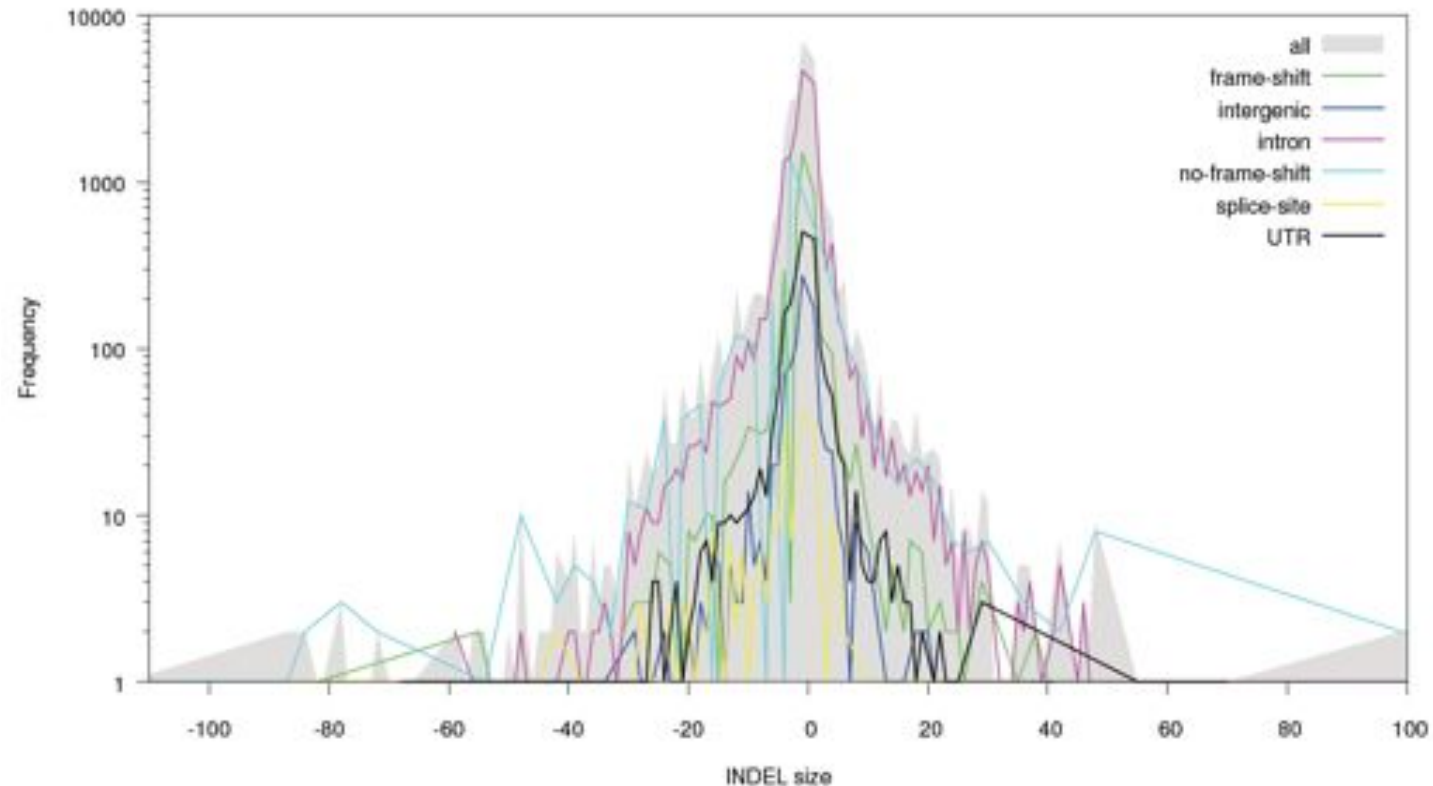
## **Many possible applications:**

- Identifying “core” genes present in all strains
- Characterizing highly variable regions (+ flanking shared)
- Cataloging sequences shared by pathogenic varieties



# Part 2: Practice

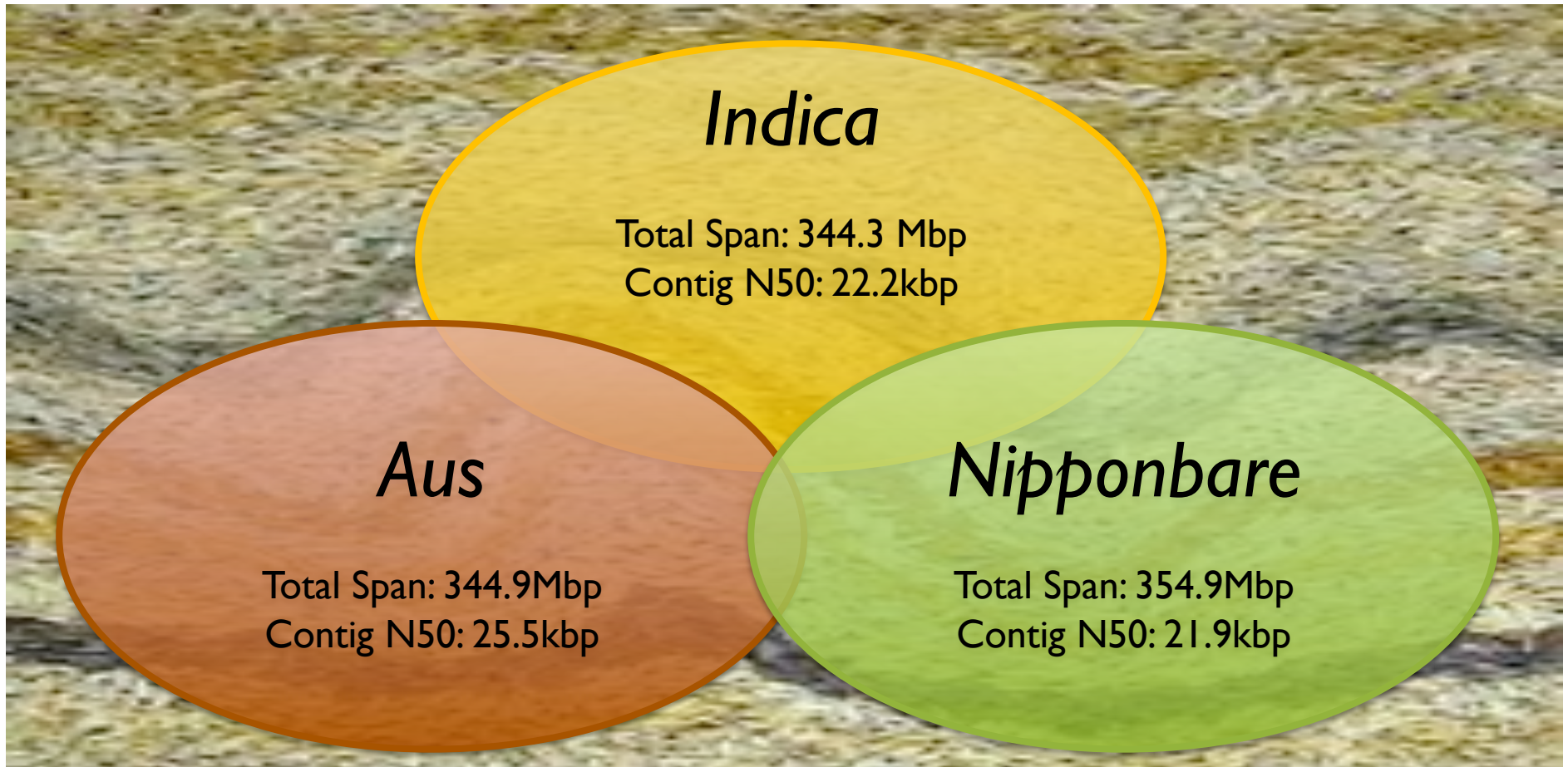
# Genetics of Autism Spectrum Disorders



1. Constructed database of  $>1M$  transmitted and de novo indels in  $\sim 1000$  families
2. For practical reasons, analysis is computed relative to the (unpatched) reference genome
  - We use population statistics to “clean” problematic regions
  - We believe we are missing and/or misinterpreting some interesting variants

**Accurate de novo and transmitted indel detection in exome-capture data using microassembly.**  
Narzisi et al. (2014) *Nature Methods*. doi:10.1038/nmeth.3069

# Population structure of *Oryza sativa*



**Whole genome de novo assemblies of three divergent strains of rice (*O. sativa*) documents novel gene space of *aus* and *indica***

Schatz, Maron, Stein et al (2014) <http://biorxiv.org/content/early/2014/04/02/003764>

# Pan-genomics of draft assemblies

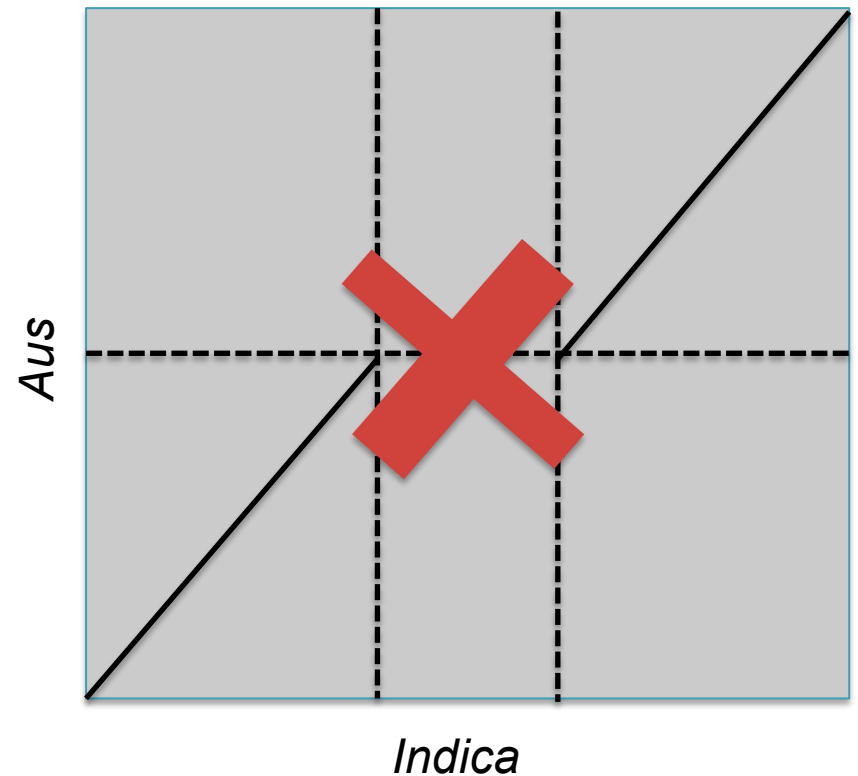
## Strategy:

1. Align the genomes to each other (MUMmer)
2. Identify segments of genome A that do not align anywhere to genome B (BEDTools)

→ Megabases specific to each genome!!!!

3. Screen regions that fail to align with their k-mer frequencies (jellyfish)
  - “Genome specific regions” averaged over 10,000x kmer coverage while unique regions were ~50x

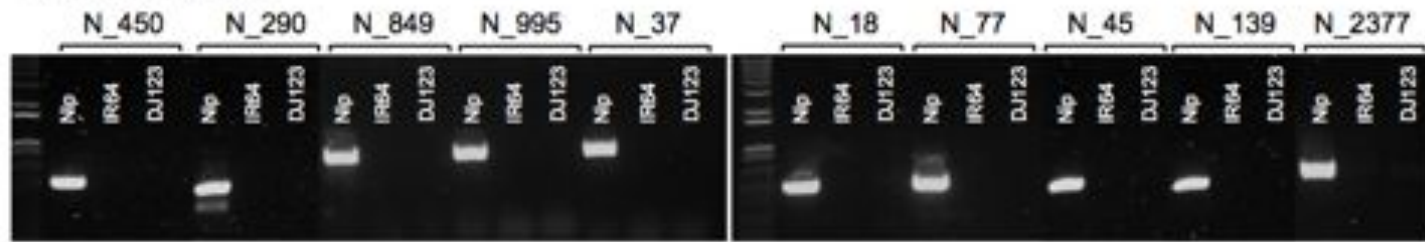
→ 100s of KB specific to each genome!!!



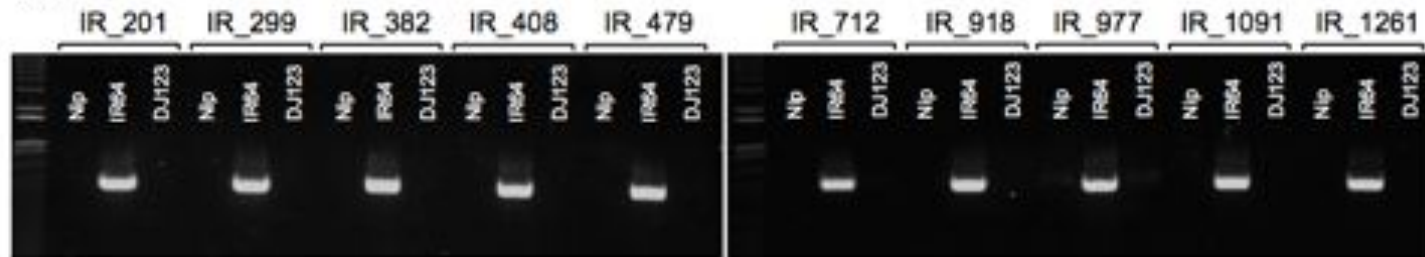


# Genome-specific Regions

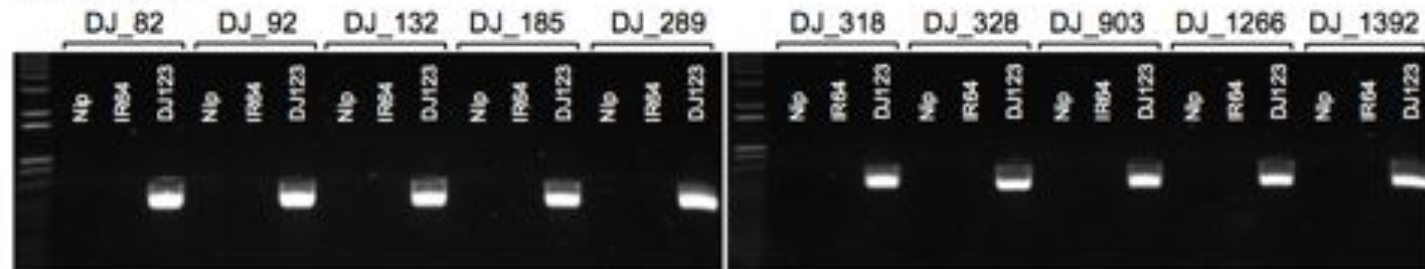
## (A) Nipponbare



## (B) IR64



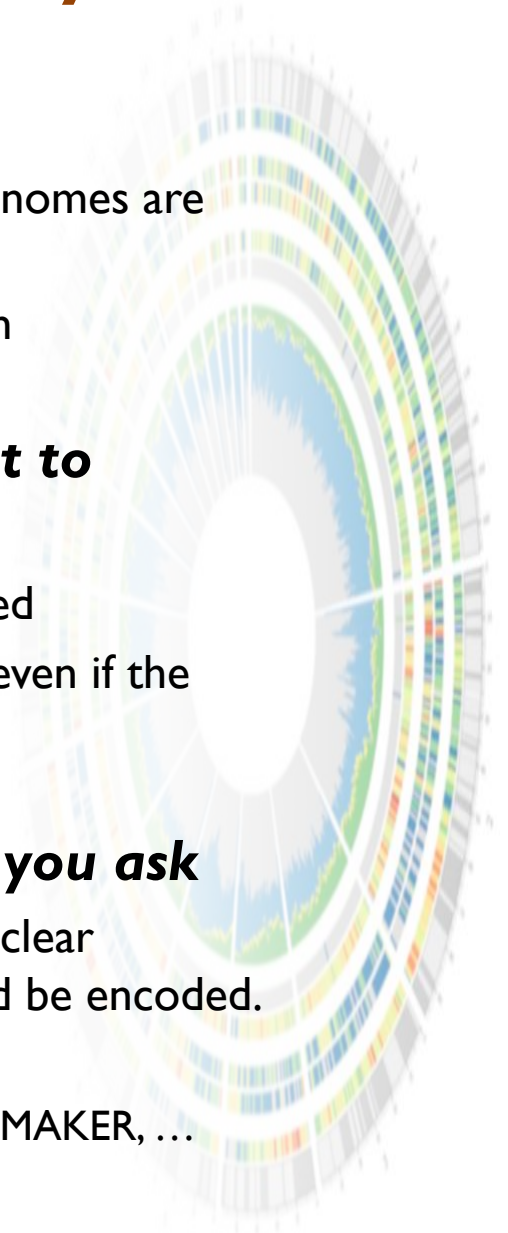
## (C) DJ123



Successfully able to identify many regions specific to each genome (30/30 PCR validation)  
Enriched for genes for disease resistance & other interesting phenotypes

# Pan-genomics Summary

- ***Now is the time to study pan-genomes***
  - Perfect assemblies of microbes and many smaller eukaryotic genomes are now routine
  - Expect to rapidly scale up these results to larger genomes soon
- ***Algorithms must scale to large collections, be robust to errors, gaps, and ambiguity***
  - Large body of assembly and alignment theory can be repurposed
  - Simple refinements, like k-mer screening, can be very effective even if the sequence is lacking
- ***The “right approach” will depend on the questions you ask***
  - We all agree we need to work from a graph, but there is not a clear consensus of what the graph should represent or how it should be encoded.
  - Ultimately the needs will be driven by applications
    - graph-BLAST, -BWA, -SAMTools, -TopHat/Cufflinks, -IGV, -UCSC, -MAKER, ...



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Levy Lab  
Lippman Lab  
Lyon Lab  
Martienssen Lab  
McCombie Lab  
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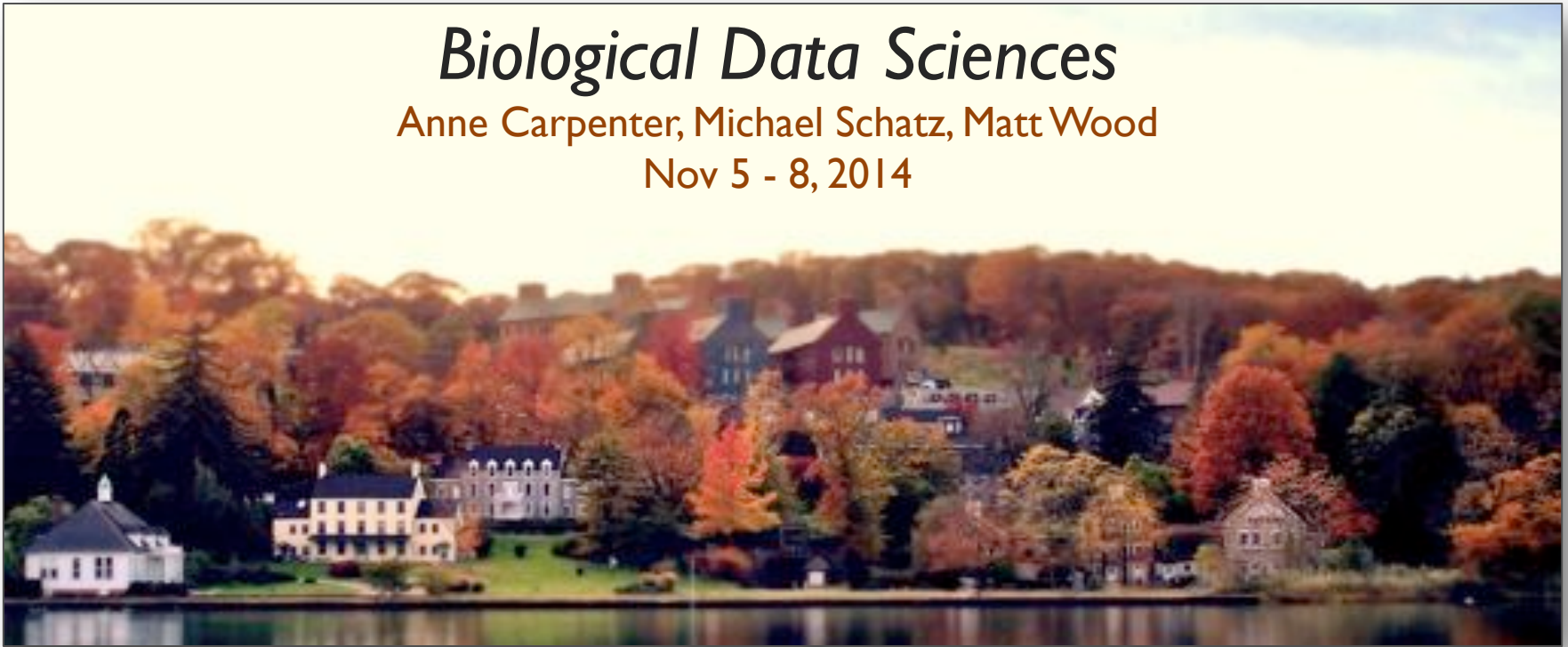
**SFARI**

SIMONS FOUNDATION  
AUTISM RESEARCH INITIATIVE

# *Biological Data Sciences*

Anne Carpenter, Michael Schatz, Matt Wood

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# Thank you

<http://schatzlab.cshl.edu>

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